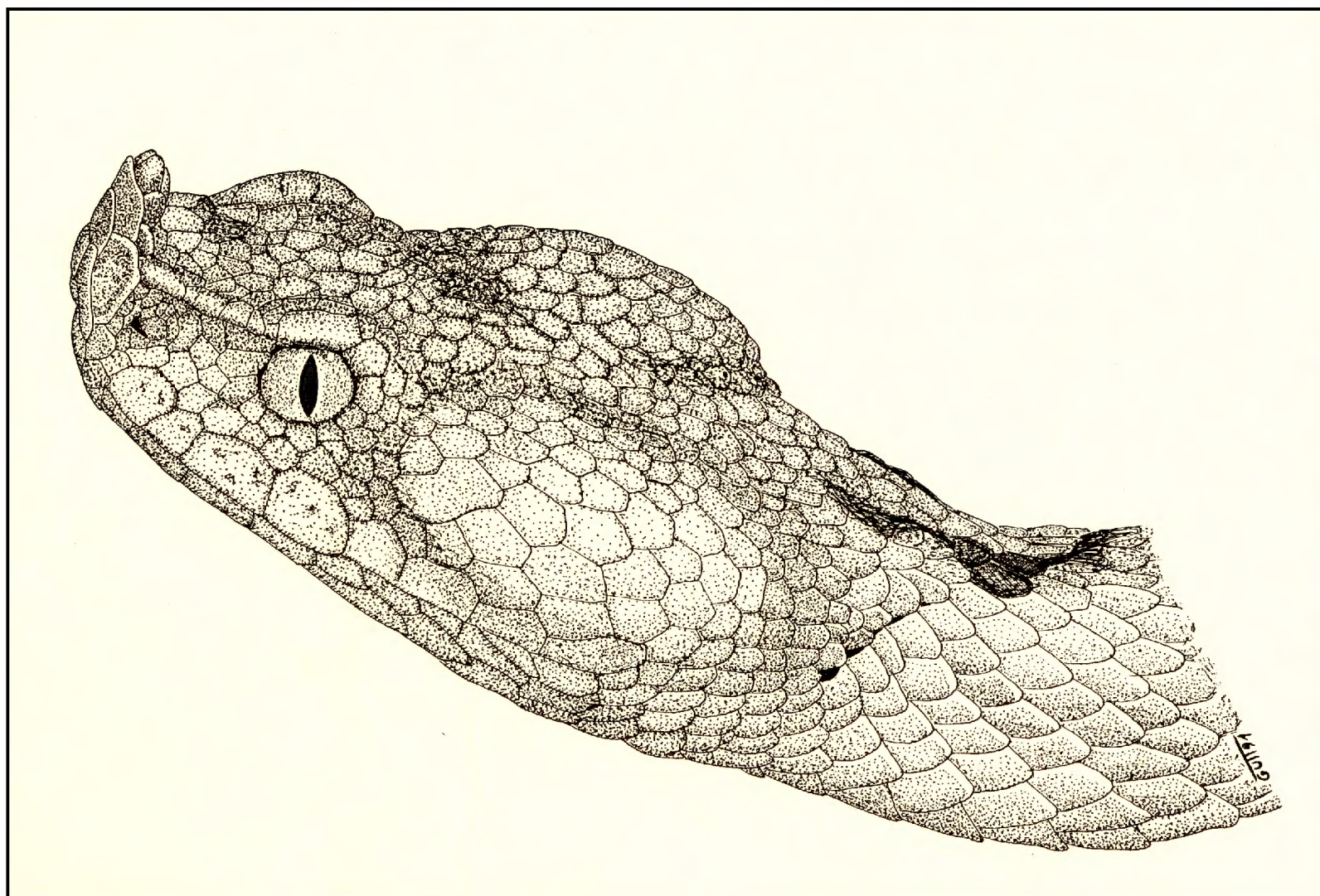

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BULLETIN OF THE CHICAGO HERPETOLOGICAL SOCIETY

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Cover: Nose-horned viper, *Vipera ammodytes*. Drawing by Thomas Guttman.

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Eastern Coachwhip (*Coluber flagellum flagellum*) Preys on Hatchling Gopher Tortoises (*Gopherus polyphemus*)

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The varied diet of the Eastern Coachwhip (*Coluber flagellum flagellum*) has been reported to include small mammals, birds and their eggs, turtles, snakes, lizards, anurans, and insects and other arthropods (Beane, 2013). Herein, I report the second documented instance of predation on the Gopher Tortoise (*Gopherus polyphemus*) by *C. f. flagellum*.

At 0940 on the morning of 7 November 2013, I found an adult female *C. f. flagellum* (snout-vent length [SVL] = 1422 mm; total length = 1829 mm; mass = 450 g [with prey]; GMNH 50649) basking near the mouth of a subadult *G. polyphemus* burrow near Mount Pleasant, Wayne County, Georgia (Figure 1). Just after capture, I noticed a pronounced bolus ca. two-thirds down the snake's body. Later the same day, the snake disgorged two hatchling-sized *G. polyphemus* (both ca. 4.8 cm in straight carapace length; GMNH 50663-50664) (Figure 2).

This is only the second report of *G. polyphemus* in the diet of *C. f. flagellum*. Douglass and Winegarner (1977) found the remains of a small *G. polyphemus* in the feces of a slightly larger *C. f. flagellum* (SVL = 1620 mm) captured at Archbold Biological Station, Highlands County, Florida. Other turtle species known to be consumed by *C. f. flagellum* include *Terrapene carolina* (Eastern Box Turtle) (Beane, 2013), *Caretta caretta* (Loggerhead Sea Turtle) (Arpayoglou and Martin, 2011), and *Kinosternon subrubrum* (Eastern Mud Turtle) (Hamilton and Pollack, 1956).



Figure 1. The adult Eastern Coachwhip that disgorged two juvenile Gopher Tortoises.

Other snakes that have been documented as predators of small Gopher Tortoises include the Eastern Indigo Snake (*Drymarchon couperi*) (Stevenson et al., 2010) and the Cottonmouth (*Agkistrodon piscivorus*) (Butler and Sowell, 1996). In fact, hatchling Gopher Tortoises are commonly eaten by Eastern Indigos (Stevenson et al., 2010). Similarly, small Texas Tortoises (*Gopherus berlandieri*) are frequently consumed by Texas Indigo Snakes (*Drymarchon melanurus erebennus*) (Werler and Dixon, 2000; Rich Kazmaier, pers. comm.).

Interestingly, there is mention in the literature of an Eastern Diamondback Rattlesnake (*Crotalus adamanteus*) eating a Gopher Tortoise, by former President Theodore Roosevelt (Roosevelt, 1917). A Captain McCann, whom Roosevelt considered “an accurate and trustworthy observer,” reported that he once found a small tortoise inside a diamondback. Roosevelt was surprised, because he assumed that diamondbacks fed exclusively on birds and mammals.

In the southeastern United States, *C. f. flagellum* attains a large size (ca. 1400–1700 mm SVL), is commonly found in sandhill habitats, and, often inhabiting *G. polyphemus* burrows, is considered a “frequent commensal” (Jackson and Milstrey, 1989). Thus, one might suspect that predation on hatchling and smaller juvenile *G. polyphemus* may occur more frequently than reflected by the published literature.

Acknowledgments

I thank Jeff Beane, Kevin Enge and Lora Smith for their help with my literature search for this note.



Figure 2. One of the juvenile Gopher Tortoises recovered from the Eastern Coachwhip (scale of ruler is in inches).

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Forest Tent Caterpillar Consumption by Wood and Mink Frogs at Lake Gegoka, Minnesota

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Introduction

Studies of dietary preferences of ranid frogs have focused on stomach contents (Krameck, 1972, Hedeén 1972, Stewart and Sandison, 1972) and competition for feeding sites (Stewart and Sandison, 1972) and have indicated that ranid frogs are dietary generalists and species' diets may be limited by competition. While caterpillars are taken by ranid frogs, Stewart and Sandison (1972) documented that both green and mink frogs consume caterpillars of noctuid and geometrid moth families. However, their results indicate that the presence of these caterpillars in mink frog diet is minor. Conversely, in Japan, the pond frog *Pelophylax nigromaculatus* (formerly *Rana nigromaculata*) may consume various lepidopteran larvae in large quantities (Hirai and Matsui, 1999; Hirai, 2002). Wood frogs are also generalist feeders (Oldfield and Moriarty, 1994), with Vogt (1981) indicating that the primary components of diet are flies and beetles.

Though caterpillars have been found in the diet of mink frogs, there has been no indication that forest tent caterpillars (*Malacosoma disstria*, Lasiocampidae) occur in their diet. These caterpillars, however, are frequent food items for various avian species, particularly northern orioles (Parry, Spence and Volney, 1997). While Meeker (1997) suggests that frogs consume forest tent caterpillars, he does not implicate any frog species. Similarly Hedeén (1972) mentions caterpillar consumption but does not specify which lepidopteran families.

Methods

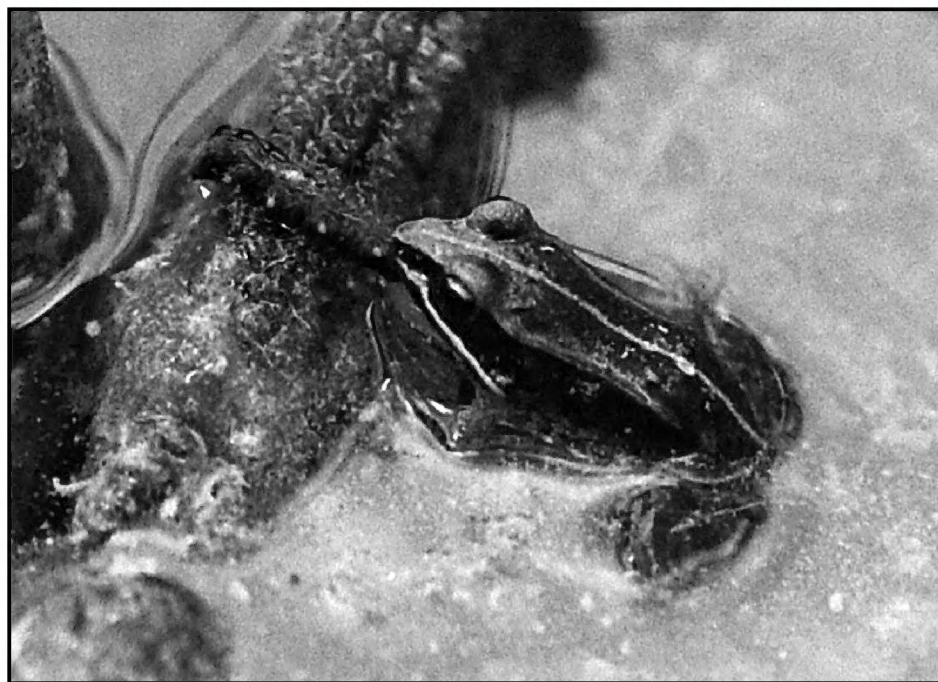
Lake Gegoka, Minnesota (47°39"N, 91°28"W) is a 1.68 × 1.2 km lake in north-central Minnesota. The study site was a 5 × 10 m bay with emergent graminoids with a shallow area overhung by red maple (*Acer rubrum*), quaking aspen (*Populus tremuloides*) and black spruce (*Picea mariana*). Several clonal

culms of emergent graminoids (probably *Juncus* sp.) made fairly linear and parallel "lanes" approximately 0.75 to 1.2 m apart making the frogs choose "lanes" in which to forage, also making observations somewhat easier. We daily collected between 20 and 30 forest tent caterpillar instars from low-hanging maple and aspen branches and from the forest floor. Between 0900 and 1100 h, I (SRJ) tossed caterpillars in a high arc to simulate the effect of a caterpillar falling from a tree, about 15 feet out from shore into shallow water and observed mink frog responses. Over the course of the experiment I altered feeding lanes daily.

Wood frogs between the shoreline and forest edge were observed feeding on naturally deposited forest tent caterpillars and also received supplemental caterpillars. We documented responses to naturally deposited and artificially introduced caterpillar proximity.

Results

Though I (SRJ) initially observed a 6-cm wood frog (*Lithobates sylvaticus*) consume a 5-cm instar tent caterpillar, wood frogs were not particularly common at the site. Those that were there tended to stay along the shoreline in the area most likely to receive fallen caterpillars. On any given day of the study only about eight to ten wood frogs were present at the study site and there was no increase in wood frog density with the supplemental feeding. We observed wood frogs among fallen tree branches at water's edge and on flattened marsh grass mats. Wood frog responses to caterpillar consumption were variable. Though the first observed wood frog successfully consumed a tent caterpillar and roughly 80% of the wood frogs were seen to successfully consume caterpillars, I (SRJ) observed that about 20% showed either a prolonged two- to three-minute open-mouthed gag response immediately following caterpillar consumption or an attack followed by immediate rejection of a caterpillar. I also



Wood frog consuming a forest tent caterpillar.



Wood frog gagging on a forest tent caterpillar.



The readily identifiable recurrent female mink frog.

observed that on several occasions wood frogs showed what appeared to be exhaustion and slowed rate of consumption when trying to eat a large instar caterpillar.

Initially, mink frogs (*Lithobates septentrionalis*) were rarely observed near the shoreline. On the first day I (SRJ) saw a single male within two meters of the shore. This individual at first paid no attention to a moving caterpillar but within five minutes approached and rapidly consumed the caterpillar and two additional caterpillars that I subsequently tossed into his vicinity.

The following morning two male mink frogs were awaiting me in the shallows and more arrived over the course of feeding including a large female with easily identified heavily dark-mottled dorsum. Over subsequent days the number of foraging mink frogs increased to a maximum of 18 to 23 individuals and by the third day of the study at least four to five frogs were positioned one meter from shore in each feeding lane in the morning. The large identifiable female hung back and came to whichever lane I conducted artificial feeding in the morning. By

the last day of feeding she was waiting for me close to shore with ten other individuals.

Discussion

Though previous studies suggest that mink frogs eat only small amounts of caterpillars, no previous study indicates that lepidopteran caterpillars were taken in large quantities or that forest tent caterpillars were taken at all. Previous studies also suggest that either through habitat preference or congeneric competition, mink frogs primarily take aquatic invertebrate prey. The results of this small study indicate that mink frogs may, if given the opportunity, consume a large volume of caterpillar prey. This work also suggests that in the absence of competitive shoreline-inhabiting ranid species, mink frogs will readily forage near the shore and occupy an area that partially overlaps the wood frog hunting zone. The behavior of the above-mentioned readily identified large female mink frog certainly echoes the finding of Krameck (1976) where frogs showed a strong tendency "to return to certain places within a hunting area." Indeed, the response of mink frogs to caterpillars suggests that in the absence of larger bullfrogs (*Lithobates catesbeianus*) and green frogs (*Lithobates clamitans*), mink frogs could forage near shore and consume large quantities of forest tent caterpillars.

Unlike the reaction of the mink frogs, wood frog numbers did not increase in response to additional caterpillar density in their shoreline to forest edge feeding zone. Additionally, the varied response of wood frogs to caterpillars suggests that these caterpillars are not a favored dietary staple. Even though some small wood frogs did take large instar caterpillars, the slowness of consumption, and the rejections suggest that many wood frogs will ignore caterpillars landing on the shoreline or the forest floor.

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Book Review: *Amphibian Conservation: Global Evidence for the Effects of Interventions*

by Rebecca K. Smith and William J. Sutherland. 2014. 276 pp.

Pelagic Publishing, Exeter, UK. ISBN 978-1-907807-86-2. Hardcover \$114.99. Paperback \$48.99.

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As readers of the *Bulletin* are aware, amphibians worldwide are facing a multitude of threats to their survival. These perils not only threaten individual amphibian populations, they threaten the very existence of many amphibian species. Since the alarm was urgently sounded in the early 1990s (e.g., Barinaga, 1990; Blaustein and Wake, 1990; Wyman, 1990), popular and scientific literature on amphibian declines has proliferated. Much of this literature is documentary in nature; that is, amphibian declines and losses are reported as they are observed. Can anything be done to stop and/or reverse these declines? The answer, per the authors of *Amphibian Conservation: Global Evidence for the Effects of Interventions* is a resounding yes!

Amphibian Conservation: Global Evidence for the Effects of Interventions is fourth in a series of books recently published by Pelagic Publishing summarizing evidence for interventions that can be taken to benefit various taxa and address particular conservation issues. The authors of *Amphibian Conservation*, Rebecca Smith and William Sutherland, both affiliated with the University of Cambridge in the United Kingdom, have previously collaborated on number 3 in the Conservation Evidence series, *Farmland Conservation: Evidence for the Effects of Interventions in Northern Europe*. Sutherland also co-authored “*Bird Conservation: Evidence for the Effects of Intervention*” and “*Bee Conservation: Evidence for the Effects of Intervention*.” A perusal of their respective University of Cambridge web pages reveals that Smith and Sutherland excel at synthesizing existing information, in addition to conducting and publishing original research.

A spectacular head-on color photograph of a masked tree-frog, *Smilisca phaeota*, taken by Robin Moore, graces the front cover. The bug-eyed frog is peering out from a hollow of a plant with hands positioned as if pulling back curtains. The image seems to capture the essence of the book—the unveiling of a compilation of conservation interventions benefitting amphibians. Readers are encouraged to visit Dr. Moore’s website (www.robindmoore.com) to learn more about him and to view more of his images.

The content of the book includes a list of individuals who comprised the nine-member advisory board, a brief (one sentence each) introduction to the authors, acknowledgements, and a six-page explanation of how to use the book followed by 14



chapters of interventions and a subject index. The “About this book” section should be consulted by every reader as it summarizes the purpose of the book, identifies the target audience, introduces the Conservation Evidence Project, names the 18 herpetological journals searched from their inception through 2012 (but neglects to name the 30 “general conservation” journals searched over the same period), names the two internet search engines used (Google Scholar and Web of Science), describes how the evidence is summarized, and defines terminology used. The target audience envisioned by the authors is “someone who has to make decisions about how best to support or conserve biodiversity.” Examples include land managers, farmers, consultants, policymakers and researchers. I think researchers, in particular, will find the voluminous literature summaries and citations especially valuable.

The goal of the book is to summarize credible actions the reader can consider to achieve on-the-ground conservation objectives for amphibians. The reader needs to be aware that several important caveats limit what sort of literature is included in the book. To be included “there must have been an intervention carried out that conservationists would do” and “the effects of the intervention must have been monitored quantitatively.” These constraints exclude studies that may have management implications but were not specifically designed to evaluate the efficacy of a particular conservation intervention. Correlative and modeling studies are among those excluded. Although these kinds of studies may not be a “smoking gun,” they can provide valuable insight into what conservation interventions may be appropriate, especially in cases where the authors’ search yielded no examples of proven interventions (see below).

Amphibian Conservation identifies 107 conservation interventions for amphibians and cites more than 400 studies from around the world. The authors do not evaluate study methodologies or conclusions; they leave judgment of the merits of each study up to the reader. Citations are not compiled at the end of the book but, rather, are provided within the text of each chapter. As a result, some papers applicable to more than one intervention are cited more than once at different locations within the book. Although the book captures a lot of literature, the authors wisely recommend that readers also conduct a systematic review of the literature for their particular situation as it “is likely to be more comprehensive than the summary of

evidence” they provide.

Amphibian Conservation interventions are organized into 14 chapters, each ranging from 1 to 54 pages in length and containing from 1 to 26 numbered subsections. Each chapter briefly introduces the threat under consideration and includes one or more key messages that provide a brief overview of the evidence. Every chapter focuses on practical interventions that can be taken to respond to the identified threat. Short (i.e., one page) chapters are those for which no evidence of intervention was found; these include: “Threat: Energy production and mining” and “Threat: Human intrusions and disturbance.” The longest chapter covers habitat restoration and creation.

To properly evaluate this book, I focused this review on four (out of many) subjects of personal interest. The first takes me back to my childhood in suburban Brookfield, Illinois. The one and only amphibian inhabiting the world of my youth was the American toad, *Anaxyrus* (a.k.a. *Bufo*) *americanus*. I encountered toads in the nearby forest preserve, in neighborhood yards and gardens, as well as in the street as they fed on insects under the light of street lamps in the evening. In addition, I often observed—with anguish—toads at the bottom of storm drains that were built into the roadbed adjacent to the curb. Because each storm drain was covered with an immovable steel grate, I was unable to extract the entrapped toads. Over time, the toads succumbed to starvation and/or desiccation. Did the authors of *Amphibian Conservation* consider this threat to amphibians and, if so, did they find a solution? Indeed, they had.

In the “Threat: Transportation and service corridors” chapter, I found a subsection entitled, “Modify gully pots and kerbs.” In the United Kingdom, the terms “gully pot” and “kerb” are synonymous with our “storm drain” and “curb,” respectively. Research in Europe found that separating storm drains from the curb by 4 inches decreased the number of great crested newts (*Triturus cristatus*) that fell in by 80%. Intrigued, I searched the internet and found the original article. Separation of the storm drain from the adjacent curb is achieved by replacing the standard section of curb with a section that has a recessed wall. Small animals following the curb wall diverge slightly into the recessed portion, thus avoiding the storm drain. This modified curb provides a simple solution to my vexing childhood dilemma of what to do about toads falling into storm drains. Perhaps urban/suburban readers of the *Bulletin* can lobby their respective community leaders to implement this solution in their neighborhoods. Interested readers are encouraged to visit www.aco.co.uk to learn more about these commercially-produced modified curbs.

As field herpetologists know, many of our more interesting amphibians cannot successfully reproduce in water bodies harboring large, predatory fishes. These fishes (sunfishes, in particular) look upon frog and salamander larvae as delectable treats. Paradoxically, I have captured larvae of fish-intolerant amphibian species co-existing with predatory fishes. How is this possible? Invariably, these larvae take refuge in aquatic vegetation.

Because a large number of people enjoy fishing, water

bodies are commonly stocked with game fish. As a result, many otherwise suitable water bodies are off limits to amphibians that are palatable to fish. Perhaps it’s possible, however, to encourage the presence of aquatic vegetation in fish ponds so they can also provide amphibian habitat. Did the authors of *Amphibian Conservation* consider this potential intervention? Yes, in Chapter 8, “Threat: Invasive alien and other problematic species.” Here, I found a subsection entitled, “Encourage aquatic plant growth as refuge against fish predation.” Unexpectedly, the authors did not find any studies demonstrating the efficacy of such an intervention. This result was surprising as I know relevant literature exists. Two studies come to mind, one conducted in the laboratory (Baber and Babbitt, 2004) and the other in the field (Shulse et al., 2012). Although both provide valuable support for providing aquatic vegetation as a means to protect amphibian larvae from fish predation—evidence that could inform real-world solutions—neither appeared to meet the criteria for inclusion in the book. The exclusion of these studies illustrates the necessity of looking beyond the literature captured by the Conservation Evidence Project (as encouraged by the authors).

One of the principal ecological disruptions in the southeastern United States coastal plain affecting amphibians is industrial pine silviculture, particularly the practice of mechanical site preparation (Means, 2005). Prior to tree planting, heavy machinery is driven over the site and the soil is roller-chopped and often disked in a fashion similar to that conducted annually on agricultural fields. On wetter landscapes, the soil is further contoured into long, raised beds on which trees are planted. Mechanical site preparation can destroy shallow, subterranean burrows and collapse entrances to larger burrows such as those of the gopher tortoise (*Gopherus polyphemus*). It also initially reduces coverage of “competing” vegetation to near zero. I wondered if the authors considered this threat and whether they found any literature that demonstrated amphibian-friendly alternatives to mechanical site preparation.

Forestry activities are addressed in Chapter 5, “Threat: Biological resource use.” A number of interventions are considered, including alternatives to removing all the trees from a site, leaving downed woody debris after harvest, and protecting riparian buffers from harvest. Mechanical site preparation, however, is not considered. Given the disruptive nature of this procedure and its suspected contribution—along with fire suppression—to the demise of several southeastern United States pineland endemic herpetofauna species (Reichling, 2008), this is unfortunate. There is a small, growing body of literature regarding the impact of pine silviculture on amphibians in the southeastern United States, but much more needs to be done.

I thought I might find alternatives to mechanical soil disturbance in chapter 2, “Agriculture.” Here, I found the promising subheading entitled, “Reduce tillage.” Alas, the authors found “no evidence for the effects of reduced tillage on amphibian populations.” Given the ever-growing human population and escalating need for food and fiber (Tilman et al., 2002), as well as the increasingly mechanized nature of food and fiber production, the potential impact of mechanized soil disruption on

amphibians seems a ripe topic for study.

Worldwide, wetland losses have been enormous (Zedler and Kercher, 2005). Wetland loss is one of the greatest threats to amphibians because so many species rely on them for reproduction. How is this addressed in *Amphibian Conservation*? Chapter 12, “Habitat restoration and creation” provides numerous examples of interventions that have been implemented to mitigate the loss of amphibian habitat, including the creation of ponds and wetlands. I was heartened to see the substantial collection of literature pertaining to this subject. Humans are very adept at tinkering with their surroundings, whether it be for human benefit (e.g., draining a marsh to increase agricultural acreage) or the benefit of wildlife (e.g., creating a wetland). The popularity and proven benefits of habitat restoration and creation is evidenced by the length of this chapter which, at 54 pages, is the longest in the book. The authors did an excellent job of providing readers with a treasure trove of information on the subject. More than any other chapter, this one gives the reader the greatest feeling of hope. The literature shows that when provided with a new home, amphibians will move in.

Amphibian Conservation is a welcome antidote to the prolif-

eration of depressing reports of amphibian declines. The book is packed with literature summaries and citations; a veritable information goldmine for graduate students and researchers. It also admirably provides decision makers with a well-researched resource of proven interventions that can be employed to stem/reverse the decline of amphibian populations. The free, downloadable pdf version and other relatively inexpensive formats that are available (eBook, for example) generously provide individuals who lack deep pockets access to this significant source of information. Interventions described in the book are available at the Conservation Evidence Project website (www.conservationevidence.com) where one can also peruse the open-access journal *Conservation Evidence*. My only caution for the reader is to take seriously the authors’ advice and perform supplementary reviews of the literature for their particular topic or species of interest. To do otherwise risks missing pertinent and important information not captured in *Amphibian Conservation*.

Acknowledgments

Mike Dloogatch kindly offered me the opportunity to conduct this review.

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What You Missed at the June Meeting

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(Photographs by Dick Buchholz)



June is always a popular meeting. It's Show and Tell. Perhaps this meeting is the essence of what your society is supposed to be. Members bring in an animal or two and talk for a bit about that animal and then answer questions. We had rare snakes and common salamanders, baby turtles and cool chameleons. Presenters ranged from the expert to the beginner, and the questions ranged from insightful to insipid. But there are always the animals and always the anticipation that precedes and curiosity that envelops every presenter. We learned about common habits of little known creatures and little known habits of common creatures. Tidbits gave us insights into how each presenter interacts with the animal being shared with us. "Pseustes" and "terrarin" mixed with "baby" and "cute." Within a limited time span we gained a better understand of not only the animals but also the reasons why so many of us devote time and energy to keeping and studying them. This meeting demonstrates the mission of your society, bringing the professional and amateur closer together in a democratic approach to herpetology. Yeah, it's that good and that much fun. Where were you?



Erica Mede talks about the diamondback terrapin (*Malaclemys terrapin*) that came through her rescue and was placed in a good home after the meeting. Adoptions are just one of the benefits of being in the society. If you're looking for a particular animal, let the adoptions chair know and you may get a nice animal while saving it from an unfortunate fate.



Mike Scott talks about his gorgeous California kingsnake (*Lampropeltis californiae*), but aren't they all? It's an easily handled animal with no inclination to bite.



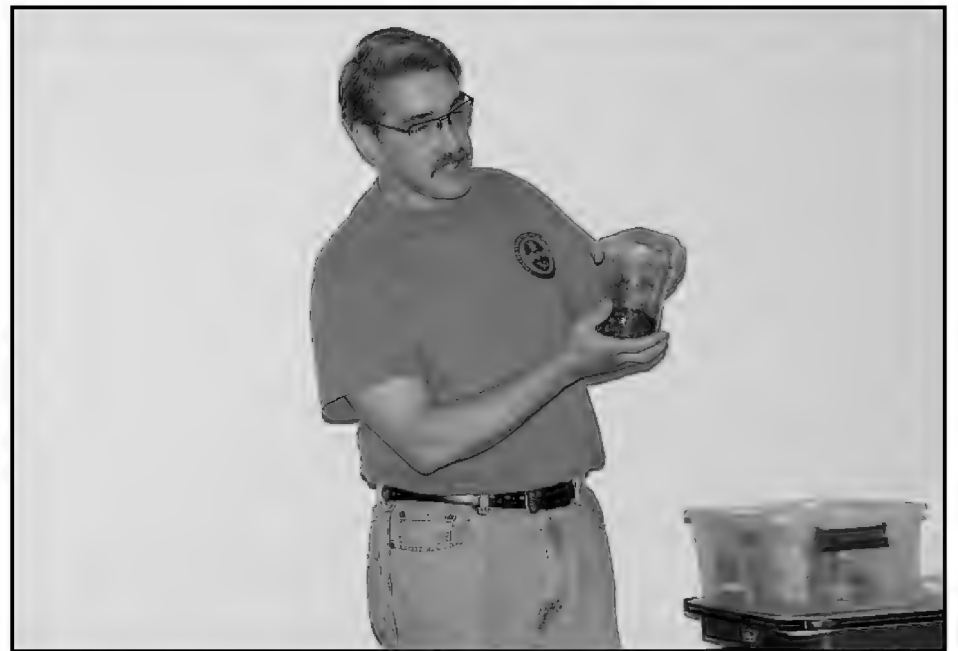
Frank Sladek brought in his red bamboo snake (*Oreocryptophis porphyraceus*). The red adds extraordinary color to its display cage, formerly a dart frog vivarium.



Marcos Glinski acquired his rainbow boa (*Epicrates cenchria*) because the species is found in his native country, Brazil. And like much of that country, his snake was gorgeous.



Joan Moore brought two of her sand boas because she was tired of hauling some of her larger snakes, but her well-fed and well cared for sand boas were pretty hefty specimens of the Indian sand boa (*Eryx johnii*), shown here, and the Kenyan sand boa (*Eryx colubrinus*).



While Rich Lamszus stretched the rules a bit (well maybe just ignored them), his talk about his spiders held our interest.



Robert Cohen showed off his gorgeous Jackson's chameleon (*Trioceros jacksonii*) while emphasizing it's a difficult species to keep. The Cohens are building a room dedicated to keeping herps.



Robert's son Noah holds an adopted green iguana (*Iguana iguana*) that should thrive in their new herp room.



Molly Carlson holds a really big ball python (*Python regius*) adopted from the many that are available through your society.



Christian holds a tiny two-day-old painted turtle (*Chrysemys picta*) he's had for two days. Many squees were heard.



Past president Jack Schoenfelder with his grandson Edward brought the only amphibian, but the tiger salamander (*Ambystoma tigrinum*) represented his class well. Jack also held his annual kids' raffle, where every kid or kid want-to-be was guaranteed a prize.



Soon to be ex-vice-president Jason Hood and wife Barbara display two of Jason's favorite snakes, a *Phrynonax* (formerly *Pseustes*) *poecilonotus* and a *Spilotes* (formerly *Pseustes*) *sulphureus*, both commonly called puffing snakes.



Kobe Taylor demonstrated an in-depth knowledge of his corn snake, including its correct scientific name of *Pantherophis guttata*.



Steve Sullivan holds aloft a Sinaloa milksnake (*Lampropeltis triangulum*), a favorite of his for shows because it's docile, attractive and encourages people to ask questions about mimicry and natural history.

Unofficial Minutes of the CHS Board Meeting, June 13, 2014

President John Archer called the meeting to order at 7:32 P.M.
Board member Andy Malawy was not present.

Officers' Reports

Recording Secretary: John Archer announced that Jenny Vollman has resigned from the office of recording secretary, and read the minutes of the May 16 board meeting. The minutes were accepted as read.

Treasurer: John Archer read the May financial report that had been e-mailed to the board Andy Malawy. An invoice from UIC for the ReptileFest facility has been received. The cost was unchanged from previous years.

Membership Secretary: Mike Dloogatch read aloud the list of lapsed members and urged board members that know any of them to remind them to renew their membership.

Corresponding Secretary: Erica Mede received and replied to an e-mail regarding red tail boas in another state. Erica will work with Jason Hood to organize the ornate box turtle field trip. The letter seeking raffle donations is ready to go.

Sergeant-at-arms: Dick Buchholz reported the number in attendance at the May general meeting was 36.

Committee Reports

Shows:

- Notebaert Nature Museum, first full weekend of each month.
- Rockin' for the Reptiles, Midwest Museum of Natural History, Sycamore, Illinois, Saturday, June 28, 12–5 P.M.
- Arlington National Laboratory picnic, July 12.
- SEWERFest, August 3.
- Bug Fest, Red Oak Nature Center, Batavia, Illinois, August 23.
- Unity Fest, Marquette Park, Chicago, September 6.
- Westmont Pet Promenade, September 13.

Junior herpers: The group will be losing four of its adult leaders. Erica Mede and Rich Lamszus were chosen as co-chairs by Jason Hood. Last month there were 40 people at the Lincoln Park Zoo field trip and 63 at the meeting. Trying to create a Field Museum behind-the-scenes field trip in October. July's speaker will be Frank Sladek.

Old business

Herp of the month: June's theme is nocturnal animals. Barbara Nieri won May.

New Illinois herp law: This law has been passed by the legislature and awaits the governor's signature.

Salamander Safari: John Archer will look into possibly holding this next spring at a DuPage County Forest Preserve, but this is still tentative.

ReptileFest 2015: The weekend of April 12–13 has been chosen. Cards need to be designed and printed. John Archer will negotiate with ZooMed about continued sponsorship.

Zoo trip: A tour for CHS members at the Columbus Zoo is

scheduled for September 20. This can only accommodate 20 people. Participants will have to travel to Columbus, Ohio, on their own and plan on at least one night in a hotel.

New Business

The board continued its discussion on the future of the CHS.

Round Table

Barbara Nieri Hood and Jason are moving to Florida next month. She is going to miss us.

Jason Hood acknowledged that we will all miss Barb more than him.

Mike Scott will have a CHS table at the next All Animal Expo.

Nancy Kloskowski had a neighbor from across the street knock on her door to ask to borrow some fuzzies.

Dick Buchholz had a great time hiking and found a stuffed coyote on the side of the road in Kentucky.

Mike Dloogatch really enjoyed the Biology of the Pitvipers conference.

Erica Mede got a legit Boba Fett helmet!

Roy Carlson went to a state park in Wisconsin and was eaten alive by gnats.

Teresa Savino ate half a pound of gnats over Memorial Day.

Jessica went to Carl Koch's funeral and will have more information on a memorial fund.

The meeting was adjourned at 9:42 P.M.

Respectfully submitted by Erica Mede



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For sale: Standard 24" Neodesha reptile cages, 3 × 6" vent on top, tempered glass front. Like new condition, no burns or stains, original glass. These cages do not have the dam (horizontal molding that crosses the entire front width of the cage, used to hold back bedding material)—easier to clean without the dam. About 20 currently available, \$45 each. Linda Malawy, (630) 717-9955, linda_malawy@hotmail.com

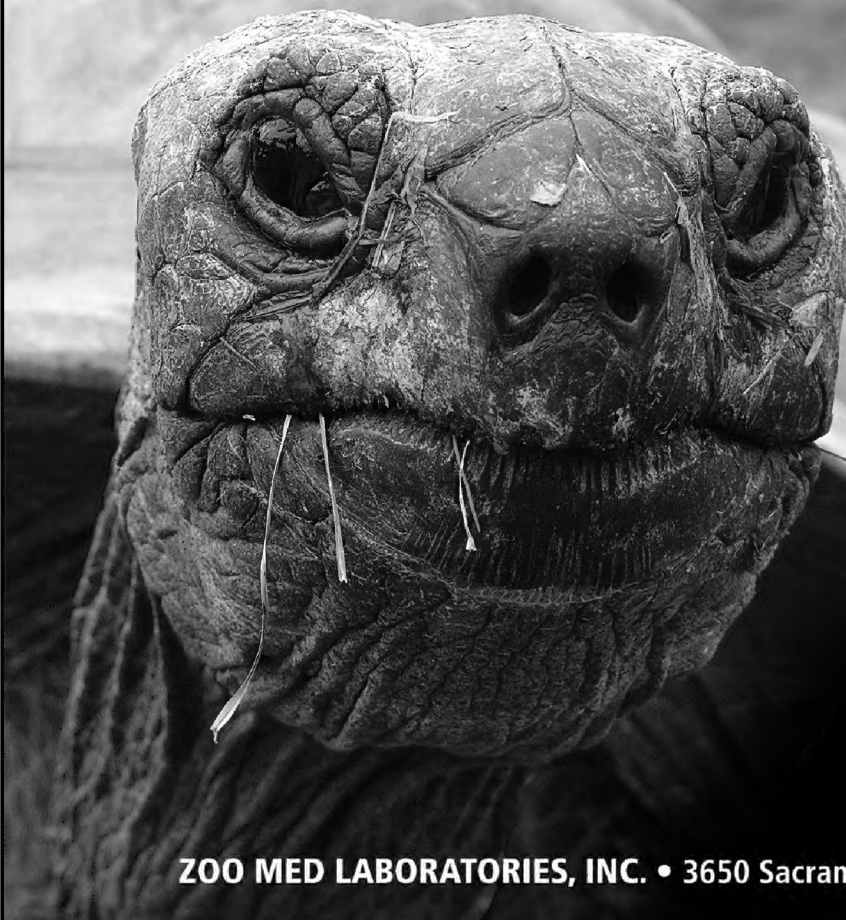
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Herp tours: **Costa Rica herping adventures**. Join a small group of fellow herpers for 7 herp-filled days. We find all types of herps, mammals, birds and insects, but our target is snakes. We average 52 per trip, and this is our 10th year doing it. If you would like to enjoy finding herps in the wild and sleep in a bed at night with air-conditioning, hot water and only unpack your suitcase once, instead of daily, then this is the place to do it. Go to our web-site <http://hiss-n-things.com> and read the highlights of our trips. Read the statistics of each trip and visit the link showing photos of the 40 different species we have found along the way. E-mail at jim.kavney@gmail.com or call Jim Kavney, 305-664-2881.

Line ads in this publication are run free for CHS members — \$2 per line for nonmembers. Any ad may be refused at the discretion of the Editor. Submit ads to mdloogatch@chicagoherp.org.

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UPCOMING MEETINGS

The next meeting of the Chicago Herpetological Society will be held at 7:30 P.M., Wednesday, July 30, at the Peggy Notebaert Nature Museum, Cannon Drive and Fullerton Parkway, in Chicago. **Dr. Robert Jadin** of Northeastern Illinois University will speak on “Untangling a Phylogenetic Knot of Venomous Snakes: Evolutionary Discoveries of Highland Pitvipers from Middle America.” From fieldwork in the tropics to morphology in the lab, this talk will shed light on how new species are found and described as well as how these advances are permitting us to make broader understandings about biogeographic patterns and treatments in modern medicine. Systematic studies like this are becoming increasingly important in our current age of rapid biodiversity loss.

A speaker has not yet been confirmed for the August 27 meeting.

The regular monthly meetings of the Chicago Herpetological Society take place at Chicago’s newest museum—the **Peggy Notebaert Nature Museum**. This beautiful building is at Fullerton Parkway and Cannon Drive, directly across Fullerton from the Lincoln Park Zoo. Meetings are held the last Wednesday of each month, from 7:30 P.M. through 9:30 P.M. Parking is free on Cannon Drive. A plethora of CTA buses stop nearby.

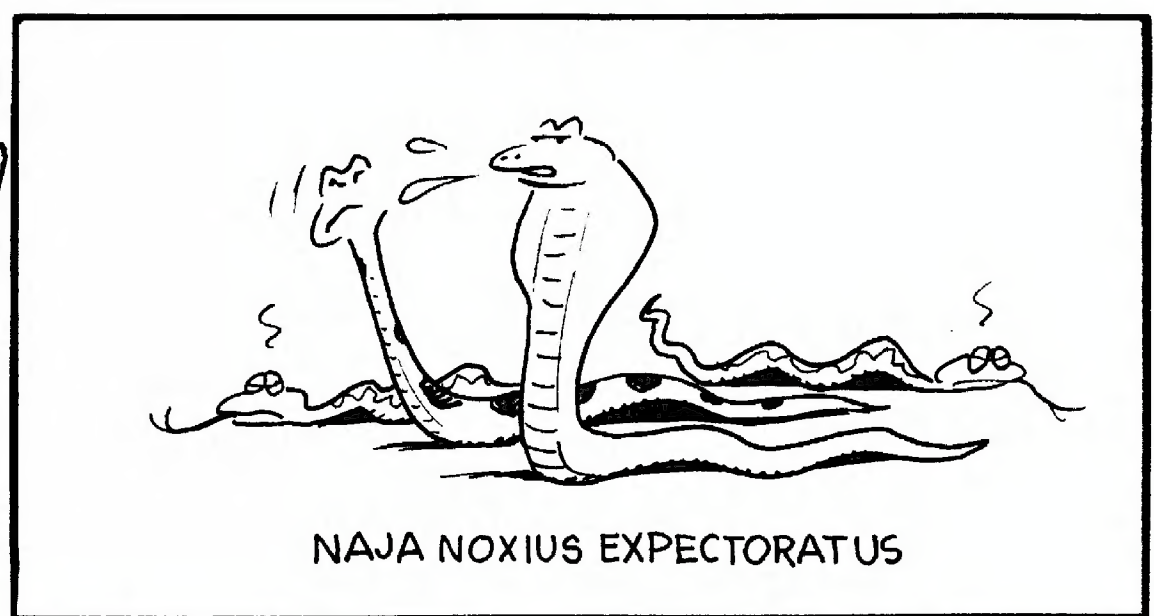
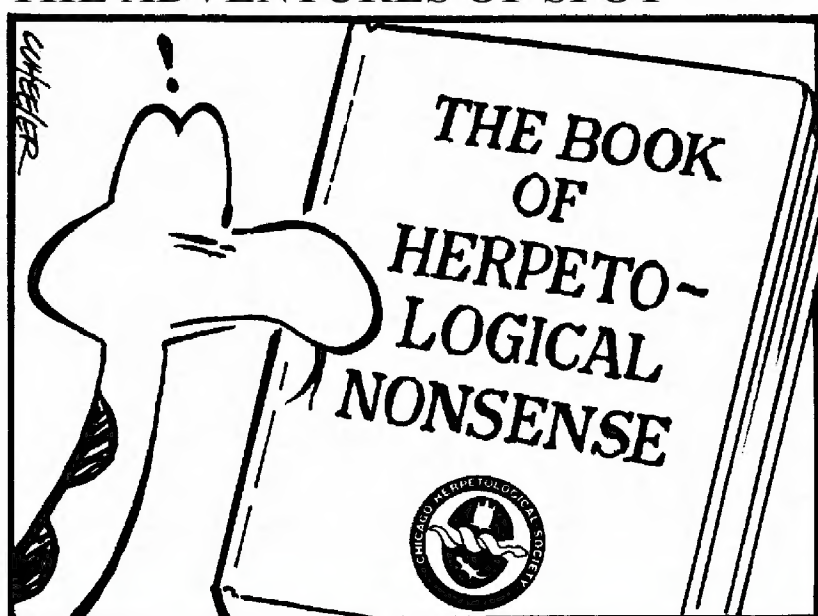
Board of Directors Meeting

Are you interested in how the decisions are made that determine how the Chicago Herpetological Society runs? And would you like to have input into those decisions? If so, mark your calendar for the next board meeting, to be held at 7:30 P.M., August 15, at the Schaumburg Township District Library, 130 S. Roselle Road, Schaumburg..

The Chicago Turtle Club

The monthly meetings of the Chicago Turtle Club are informal; questions, children and animals are welcome. Meetings normally take place at the North Park Village Nature Center, 5801 N. Pulaski, in Chicago. Parking is free. For more info visit the group’s Facebook page.

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